

Review

Cannabinoids and the gastrointestinal tract

Summary

The enteric nervous system of several species, including the mouse, rat, guinea pig and humans, contains cannabinoid CB₁ receptors that depress gastrointestinal motility, mainly by inhibiting ongoing contractile transmitter release. Signs of this depressant effect are, in the whole organism, delayed gastric emptying and inhibition of the transit of non-absorbable markers through the small intestine and, in isolated strips of ileal tissue, inhibition of evoked acetylcholine release, peristalsis, and cholinergic and non-adrenergic non-cholinergic (NANC) contractions of longitudinal or circular smooth muscle. These are contractions evoked electrically or by agents that are thought to stimulate contractile transmitter release either in tissue taken from morphine pretreated animals (naloxone) or in unpretreated tissue (γ -aminobutyric acid and 5-hydroxytryptamine). The inhibitory effects of cannabinoid receptor agonists on gastric emptying and intestinal transit are mediated to some extent by CB₁ receptors in the brain as well as by enteric CB₁ receptors. Gastric acid secretion is also inhibited in response to CB₁ receptor activation, although the detailed underlying mechanism has yet to be elucidated. Cannabinoid receptor agonists delay gastric emptying in humans as well as in rodents and probably also inhibit human gastric acid secretion. Cannabinoid pretreatment induces tolerance to the inhibitory effects of cannabinoid receptor agonists on gastrointestinal motility. Findings that the CB₁ selective antagonist/inverse agonist SR141716A produces *in vivo* and *in vitro* signs of increased motility of rodent small intestine probably reflect the presence in the enteric nervous system of a population of CB₁ receptors that are precoupled to their effector mechanisms. SR141716A has been reported not to behave in this manner in the myenteric plexus-longitudinal muscle preparation (MPLM) of human ileum unless this has first been rendered cannabinoid tolerant. Nor has it been found to induce “withdrawal” contractions in cannabinoid tolerant guinea pig ileal MPLM. Further research is required to investigate the role both of endogenous cannabinoid receptor agonists and of non-CB₁ cannabinoid receptors in the gastrointestinal tract. The extent to which the effects on gastrointestinal function of cannabinoid receptor agonists or antagonists/inverse agonists can be exploited therapeutically has yet to be investigated as has the extent to which these drugs can provoke unwanted effects in the gastrointestinal tract when used for other therapeutic purposes.

The endocannabinoid system

The plant *Cannabis sativa* is the source of a set of more than 60 oxygen containing aromatic hydrocarbon compounds called cannabinoids, of which Δ^9 -tetrahydrocannabinol (Δ^9 -THC) is the main psychotropic constituent. Of the other plant cannabinoids, those which have been most investigated are Δ^8 -THC which has similar pharmacological properties to Δ^9 -THC, cannabidiol, which has much weaker psychotropic properties than Δ^9 -THC, and cannabidiol, which lacks psychotropic activity.¹ The effects of Δ^9 -THC are mediated primarily by cannabinoid receptors, at least two types of which are present in mammalian tissues. These are CB₁ and CB₂ receptors and both are members of the superfamily of G protein coupled

receptors.² CB₁ receptors are found mainly on neurones in the brain, spinal cord, and peripheral nervous system, one of their functions being to modulate neurotransmitter release. The physiological roles of CB₂ receptors, which are expressed mainly by immune cells, are proving more difficult to establish. Endogenous ligands for cannabinoid receptors have been identified, the most important being anandamide (arachidonyl ethanolamide) and 2-arachidonyl glycerol (2-AG). There is evidence that anandamide, and possibly also 2-AG, are removed from the extracellular space by a carrier mediated uptake process that is present in neurones and astrocytes.^{3,4} Once within the cell, anandamide is hydrolysed to arachidonic acid and ethanolamine by the enzyme fatty acid amide hydrolase (FAAH).³ This microsomal enzyme, which is found both in neurones and in some non-neuronal tissues, can also catalyse the hydrolysis of 2-AG. The “endocannabinoids”, anandamide and 2-AG, and their receptors constitute the “endocannabinoid system”.

As detailed elsewhere,^{5,6} a wide range of ligands for CB₁ and CB₂ receptors have now been developed. Most notable among the CB₁ selective ligands are the agonists (*R*)-(+)-arachidonyl-1'-hydroxy-2'-propylamide (methanandamide), arachidonyl-2'-chloroethylamide, and arachidonyl-cyclopropylamide, and the antagonists/inverse agonists SR141716A and LY320135. Of the CB₂ selective agonists, methanandamide is less susceptible than anandamide to hydrolysis by FAAH. Important CB₂ selective ligands include the agonists L759633, L759656, JWH-133, and HU-308, and the antagonist/inverse agonist SR144528. Inhibitors of endocannabinoid uptake and metabolism are also now available.^{5,6} Among these are the anandamide uptake inhibitor *N*-(4-hydroxyphenyl) arachidonylamide (AM404), and the potent FAAH inhibitors palmitylsulphonyl fluoride (AM374) and stearylsulphonyl fluoride (AM381). Many of the experiments described in this review have been performed with cannabinoid receptor agonists that have similar affinities for CB₁ and CB₂ receptors. Of these, the most commonly used have been WIN55212, which exhibits marginal CB₂ selectivity, the “classical” cannabinoid Δ^9 -THC, and the “non-classical” cannabinoid CP55940.^{5,6} These agonists contain chiral centres and show marked stereoselectivity in both binding and functional assays. For classical and non-classical cannabinoids, those with the same absolute stereochemistry as (–)- Δ^9 -THC at 6a and 10a (6a*R*, 10a*R*) have the greater activity (the (–)-enantiomers). However, for WIN55212, the *R*-(+) enantiomer is the more active.

There is now good evidence for the presence of CB₁ and CB₂ receptors in the gastrointestinal tract. This article summarises this evidence and also considers what is currently known about the precise location of these receptors and the effects they mediate.

Abbreviations used in this paper: MPLM, myenteric plexus-longitudinal muscle preparation; Δ^9 -THC, Δ^9 -tetrahydrocannabinol; 2-AG, 2-arachidonyl glycerol; FAAH, fatty acid amide hydrolase; AM404, *N*-(4-hydroxyphenyl) arachidonylamide; L-NAME, *N*^G-nitro-L-arginine methyl ester; AER, ascending enteric reflex; NANC, non-adrenergic non-cholinergic; PMSF, phenylmethylsulphonyl fluoride.

Cannabinoids inhibit electrically evoked contractions of isolated small intestine

The ability of cannabinoids to inhibit electrically evoked contractions of isolated preparations of small intestine mounted in organ baths and the underlying mechanisms have been the subject of many investigations over the past 30 years. These have involved experiments, mainly with guinea pig tissue, in which contractions have been produced by electrical stimulation of prejunctional neurones rather than by direct stimulation of intestinal smooth muscle.

Among the first of these experiments were those described by Gill and colleagues.^{7,8} They investigated the response of the guinea pig isolated ileum to Δ^9 -THC or to various subfractions of petrol soluble and petrol insoluble fractions of tincture of cannabis BPC, then still a licensed medicine in the UK. The petrol insoluble fraction was found to contain material with atropine-like properties that opposed ileal contractions which had been evoked either by electrical stimulation or by acetylcholine. It also contained two substances with muscarinic properties that induced contractions of resting ileum. Two subfractions of the petrol soluble fraction also proved to have pharmacological activity. These were an ether eluate (fraction III) and one component of an ether/petroleum spirit eluate that contained Δ^9 -THC (fraction IIc). The effects of fraction III resembled the non-specific effect of alcohols: depression of both electrically evoked and acetylcholine evoked contractions. On the other hand, fraction IIc depressed the twitch response to electrical stimulation without affecting the response to added acetylcholine. This was most likely due to the Δ^9 -THC present in this subfraction as similar results were obtained with pure Δ^9 -THC. This effect of Δ^9 -THC has since been confirmed many times using isolated strips of whole ileum or of ileal longitudinal or circular muscle together with the neuronal networks that contain the final motor neurones that regulate the activity of smooth muscle. Experiments with such preparations have provided conclusive evidence that the small intestine contains CB₁ receptors and that cannabinoid induced inhibition of electrically evoked contractions of small intestine is mediated by these receptors. This evidence is summarised below.

- Cannabinoid receptor agonists show high potency and remarkable stereoselectivity as inhibitors of electrically evoked contractions of isolated whole ileum or of MPLM of the ileum (table 1).
- As is to be expected for a receptor mediated response, the ability of particular cannabinoid receptor agonists to inhibit electrically evoked contractions of preparations

of human or guinea pig ileum is concentration dependent and the relationship between log concentration and response is sigmoid in nature.^{9,10,12,13,17,18}

- The rank order of potencies of cannabinoids for inhibition of electrically evoked contractions of guinea pig MPLM or whole ileum correlates well with that of their psychotropic potencies and of their affinities for specific CB₁ binding sites in brain tissue^{10,12-14,19} (see also Pertwee³).
- Using a quantitative autoradiographic binding technique, Lynn and Herkenham²⁰ demonstrated that Peyer's patches from rat intestinal tract contain high affinity specific binding sites for [³H]CP55940 and that the affinities of a selected range of cannabinoids for these binding sites correspond reasonably closely to their affinities for [³H]CP55940 binding sites of rat brain. They also found [³H]CP55940 to be more potently displaced from its Peyer's patch binding sites by (-)- Δ^9 -THC or CP55940 than by the (+)-enantiomers of these cannabinoids or by cannabidiol, each of which has relatively low cannabinoid receptor affinity. Although Lynn and Herkenham²⁰ were able to visualise [³H]CP55940 binding sites in Peyer's patches located in rat jejunum, ileum, and rectum by autoradiography, they did not observe such binding sites elsewhere in these regions of the intestine or indeed in the rat stomach, duodenum, caecum, or colon. These negative findings may be an indication that cannabinoid receptors in the gastrointestinal tract are localised in discrete regions such as nerve terminals that form only a small part of the total tissue mass. Thus using the more sensitive technique of immunohistochemistry, it has been possible to visualise CB₁ receptor immunoreactivity on neurones both in rat and guinea pig small intestine²¹ and in rat embryo digestive tract.²² In addition, saturation and displacement binding assays with membrane fractions obtained from guinea pig MPLM homogenates have revealed the presence of specific binding sites with properties similar to those of guinea pig brain CB₁ receptors.²³
- Polymerase chain reaction performed on cDNA prepared from RNA isolated from whole small intestine of guinea pig has revealed the presence in this tissue of both CB₁ and CB₂ or CB₂-like RNA.²⁴ However, CB₁ but not CB₂ or CB₂-like mRNA was detected in guinea pig MPLM. CB₁ mRNA has also been detected in human stomach and colon²⁵ and all along the digestive tract of rat embryo.²²
- Cannabinoid receptor agonist induced inhibition of electrically evoked contractions of guinea pig or human

Table 1 Stereoselective effects of cannabinoids on electrically evoked contractions of guinea pig isolated ileum or of the myenteric plexus-longitudinal muscle preparation (MPLM) of guinea pig ileum

Procedure	Cannabinoid	Concentration	Effect on twitch amplitude	Ref
Supramaximal stimulation of whole ileum at 0.1 Hz (ethanol)	(-)- Δ^8 -THC	79.5 nM	Inhibition	9
	(+)- Δ^8 -THC	318nM	None	
	(-)-11-OH- Δ^9 -THC	ca. 3 nM upwards	Inhibition	
	(+)-11-OH- Δ^9 -THC	61 nM	None	
Stimulation of whole ileum at 0.1 Hz (ethanol)	(-)- Δ^9 -THC	*EC ₅₀ =100 nM	Inhibition	10
	(+)- Δ^9 -THC	2 μ M	None	
Supramaximal stimulation of MPLM at 0.2 Hz (Cremophor EL)	(-)- Δ^9 -THC	*EC ₅₀ =125 nM	Inhibition	11
	(+)- Δ^9 -THC	*EC ₅₀ =3.1 μ M	Inhibition	
Supramaximal stimulation of MPLM at 0.1 Hz (Tween 80)	HU-210	*EC ₅₀ =1.4 nM	Inhibition	12
	HU-211	*EC ₅₀ >316 nM	Inhibition (316 nM)	
Supramaximal stimulation of MPLM at 0.1 Hz (Tween 80)	CP55940†	*EC ₅₀ =3.5 nM	Inhibition	13
	CP56667	*EC ₅₀ =162.9 nM	Inhibition	
Supramaximal stimulation of MPLM at 0.1 Hz (Tween 80)	(+)-WIN55212	EC ₅₀ =5.5 nM	Inhibition	13 14
	(-)-WIN55212	1 μ M	Negligible	

*Concentration producing 50% inhibition of the twitch response. Cannabinoid vehicle used is shown in parentheses.

†CP55940 has also been found to be more potent than CP56667 as a twitch inhibitor of mouse MPLM.¹⁵

CP56667 is the (+)-enantiomer of CP55940 and HU-211 is the (+)-enantiomer of HU-210 (dimethylheptyl-11-hydroxy- Δ^8 -tetrahydrocannabinol).

The samples of (+)- Δ^8 -THC and (+)- Δ^9 -THC used probably contained 5–10% of the corresponding (-)-enantiomer.¹⁶

- MPLM or of strips of circular smooth muscle of guinea pig ileum are prevented or reversed by SR141716A at concentrations well below 1 μM but not by SR144528.^{13 14 18 26} Dissociation constant values obtained for SR141716A against (+)-WIN55212 or CP55940 correspond closely to the dissociation constant of SR141716A for displacement of [³H]CP55940 from specific binding sites on guinea pig brain membranes²³ or from human CB₁ receptors.²⁷
- CB₁ receptors are negatively coupled through G_{i/o} proteins to adenylate cyclase and calcium channels.² Consequently, the observations that (+)-WIN55212 induced inhibition of electrically evoked contractions of guinea pig MPLM can be enhanced by lowering the external calcium concentration and attenuated by elevating the external calcium concentration or by exposing the tissue to forskolin, 8-bromo-cAMP, or the phosphodiesterase inhibitor 3-isobutyl-1-methylxanthine, provide further evidence that this inhibitory effect is mediated by CB₁ receptors.²⁸
 - Certain types of non-cannabinoid receptors can also mediate inhibition of electrically evoked twitches of guinea pig MPLM. As these include opioid and α_2 adrenergic receptors, it is noteworthy that selective antagonists of one or other of these receptor types do not block or reverse the inhibitory effect of cannabinoid receptor agonists in this tissue.¹²⁻¹⁴ It has also been found that (+)-WIN55212 induced inhibition of electrically evoked contractions of circular smooth muscle of guinea pig ileum is unaffected by naloxone or by the nitric oxide synthase inhibitor, N^G-nitro-L-arginine methyl ester (L-NAME).²⁶

Enteric CB₁ receptors mediate inhibition of evoked acetylcholine release

Evidence that CB₁ receptors mediate inhibition of electrically evoked contractions of small intestine or MPLM by decreasing evoked release of acetylcholine can be summarised as follows.

- Concentrations of a cannabis extract ("fraction IIc") or of Δ^9 -THC, 11-hydroxy- Δ^8 -THC, (+)-WIN55212, CP55940, or anandamide that inhibit electrically evoked contractions of guinea pig whole ileum or preparations of longitudinal (MPLM) or circular smooth muscle of guinea pig or human ileum have been found not to reduce the contractile response of these preparations to added acetylcholine,^{7-11 13 14 26 29} carbachol,¹⁸ substance P, or histamine,^{9 26 29} each of which acts directly on smooth muscle. On the other hand, 11-hydroxy- Δ^8 -THC, Δ^9 -THC, and L-nantradol have been shown to reduce the contractile response of guinea pig whole ileum or MPLM to 5-hydroxytryptamine or γ -aminobutyric acid, drugs that act prejunctionally to increase acetylcholine release.^{9 10 29-31} There is also a report that the ability of 11-hydroxy- Δ^9 -THC to inhibit electrically evoked contractions of strips of guinea pig ileum can be blocked by enhancing extracellular concentrations of acetylcholine by inhibiting acetylcholinesterase with physostigmine.⁹ It is noteworthy however that Layman and Milton³² found that submicromolar concentrations of Δ^9 -THC inhibited both electrically induced contractions of guinea pig ileum and contractions induced by administration of acetylcholine or histamine. These effects of Δ^9 -THC on acetylcholine and histamine induced contractions were probably non-specific in nature and unrelated to inhibition of the twitch response as cannabidiol was no less potent than Δ^9 -THC in inhibiting the responses to acetylcholine and histamine but did not inhibit the electrically evoked contractions.

- Experiments with guinea pig ileum or MPLM have shown Δ^9 -THC to decrease the output of acetylcholine from unstimulated tissues.^{32 33} The effect of Δ^9 -THC was greatest when the initial spontaneous release of acetylcholine was high, suggesting that the cannabinoid was acting to reduce acetylcholine output to a basal level.³³ This hypothesis is supported by the more recent finding that some concentrations of (+)-WIN55212 and CP55940 that inhibit electrically evoked contractions of guinea pig MPLM also inhibit electrically evoked acetylcholine release.^{13 14} There is also a brief report that acetylcholine release from guinea pig MPLM evoked by electrical stimulation or by added reseriferatoxin or 5-hydroxytryptamine can be decreased by anandamide.³⁴ Unexpectedly, although cannabidiol has generally been found not to inhibit electrically evoked contractions of guinea pig ileum,^{9 10 29 32} there is one report that cannabidiol can reduce spontaneous acetylcholine output from strips of guinea pig ileum and that it elicits this response with greater potency than Δ^9 -THC.³²
- As is to be expected for a CB₁ receptor mediated response, the ability of (+)-WIN55212 and CP55940 to inhibit electrically evoked release of acetylcholine from guinea pig MPLM is not shared by (-)-WIN55212 and can be prevented or reversed by SR141716A.^{13 14} The dissociation constant of SR141716A for antagonism of (+)-WIN55212 is not far removed from its dissociation constant for displacement of [³H]CP55940 from guinea pig brain or human CB₁ receptors.^{23 27}

CB₁ receptors in guinea pig MPLM are located prejunctionally

There is good evidence that the CB₁ receptors that apparently mediate inhibition of electrically evoked contractions of guinea pig MPLM are located prejunctionally on a subset of myenteric neurones of guinea pig small intestine that serve as the final motor neurones to longitudinal muscles. This is summarised below.

- The cannabinoid receptor agonists (+)-WIN55212 and CP55940 can potently reduce the amplitudes of evoked fast excitatory synaptic potentials of myenteric neurones of the S cell type, measured using conventional intracellular recording techniques.³⁵ This inhibitory effect on cholinergic synaptic transmission was found to be dose related and stereoselective and to be reversed by SR141716A (1 μM). Some S neurones were insensitive to (+)-WIN55212, suggesting that not all such neurones bear cannabinoid receptors. (+)-WIN55212 also reduced the amplitude of slow (NANC) excitatory synaptic potentials of myenteric neurones of the S cell type evoked by repetitive focal stimulation of interganglionic nerve fibres.
- Buckley and colleagues²² have reported the presence of CB₁ mRNA in the myenteric and submucosal plexus of rat embryo digestive tract. In experiments with an affinity purified polyclonal antibody directed against the N terminal residues of the rat cloned CB₁ receptor they also detected CB₁ immunostaining at these sites. Using the same antibody to visualise the cellular distribution of CB₁ receptors in adult guinea pig and rat MPLM by scanning confocal microscopy, Anavi-Goffer and colleagues³¹ have confirmed the presence of CB₁ receptors on neurones of the myenteric plexus. Their results also support a close association of the CB₁ receptors with cholinergic neurones of the myenteric plexus. In the guinea pig, punctate CB₁ labelling was observed on fine nerve fibres within myenteric ganglia. CB₁ positive fibres were seen to surround choline acetyltransferase containing ganglionic soma that were presumably the cell bodies of cholinergic neurones. CB₁ receptor staining in

guinea pig MPLM was also closely associated with glutamic acid decarboxylase containing fibres and hence presumably with GABAergic fibres. In rat MPLM, punctate labelling was observed within myenteric ganglia only on cell bodies. The CB₁ receptor staining was associated with choline acetyltransferase containing ganglionic soma, indicating rat CB₁ to be expressed by some cholinergic neurones. However, GABAergic fibres seemed to be completely dissociated from CB₁ receptor labelling. In MPLM of both species, CB₁ receptor staining was associated with synaptic labelling.

- Electrically evoked contractions of guinea pig and human MPLM that are susceptible to inhibition by cannabinoid receptor agonists can also be more or less completely abolished by 0.1, 0.2, or 1 μ M tetrodotoxin, a specific Na⁺ channel blocker.^{18, 36} This finding implies that cannabinoids inhibit contractions caused by contractile transmitter release from neurones rather than by direct electrical stimulation of intestinal smooth muscle. It is therefore at least consistent with the hypothesis that cannabinoids inhibit electrically evoked contractions of MPLM by acting at prejunctional sites.
- The presence of CB₁ receptors on intestinal prejunctional neurones is also supported by the evidence that CB₁ receptors in the small intestine mediate inhibition of evoked neuronal release of acetylcholine (see above).

Cannabinoid receptor agonists inhibit peristalsis in guinea pig isolated ileum

Izzo and colleagues³⁷ have recently demonstrated the effects of (+)-WIN55212 and CP55940 on peristalsis induced in segments of guinea pig isolated ileum by continuous intraluminal infusion of Krebs solution. These cannabinoids affected both the initial "preparatory phase" of peristalsis, in which the longitudinal muscle contracts in response to infusion of fluid, and the subsequent "emptying phase", in which the circular muscle contracts towards the aboral end of the intestine in a wave-like manner. More specifically, WIN55212 and CP55940 increased threshold pressure and volume for triggering peristalsis and decreased both longitudinal muscle reflex contraction occurring during the preparatory phase and resistance of the intestinal wall to the infused liquid (compliance), measured at the end of this phase. They also decreased maximal ejection pressure, measured during the emptying phase. All of these effects of (+)-WIN55212 and CP55940 were completely counteracted by 100 nM SR141716A but not by 100 nM SR144528, suggesting they were CB₁ mediated.

Similarly, Heinemann and colleagues³⁸ have reported that methanandamide can produce a concentration related inhibition of distension induced propulsive motility of lumenally perfused strips of guinea pig isolated ileum. This effect could be attenuated by SR141716A (1 μ M), by the nitric oxide synthase inhibitor L-NAME, and by apamine, an inhibitor of small conductance calcium dependent potassium channels that in guinea pig small intestine are thought to mediate fast neuromuscular transmission from inhibitory motor neurones. However, it was not affected by naloxone or by the P2 ATP receptor antagonists suramin and pyridoxal-phosphate-6-azophenyl-2',4'-disulphonic acid. Heinemann and colleagues³⁸ also found that ascending enteric reflex (AER) contraction of the circular muscle of the ileum induced by inflation of an intraluminal balloon could be inhibited by methanandamide, again in a SR141716A sensitive manner. Their results led Heinemann and colleagues³⁸ to postulate that the effect of methanandamide on propulsive peristalsis of guinea pig isolated ileum is mediated by CB₁ receptors which act (i) to activate inhibitory enteric motor pathways that oppose distension induced peristalsis by causing release of nitric

oxide and apamine sensitive transmitters and (ii) to inhibit excitatory enteric motor pathways that mediate AER contraction of circular muscle in response to distension.

Heinemann and colleagues³⁸ also found that under conditions in which peristalsis is thought to be maintained by endogenous tachykinins (blockade of cholinergic transmission with atropine or hexamethonium plus restoration of peristalsis with naloxone), the ability of methanandamide to inhibit propulsive peristalsis and AER contractions was not only preserved but enhanced. This finding prompted the proposal that, when activated, CB₁ receptors of guinea pig ileum suppress propulsive peristalsis and AER contractions by inhibiting both cholinergic and non-cholinergic transmission. Evidence for this hypothesis has also been obtained by Izzo and colleagues²⁶ in experiments with a circular muscle preparation. They found (+)-WIN55212 to inhibit cholinergic and NANC contractions evoked in this preparation by electrical stimulation and that both these inhibitory effects were potently antagonised by SR141716A. Similar results were obtained with anandamide. Interestingly, apamin reduced the inhibitory effect of (+)-WIN55212 on cholinergic contractions, pointing to cannabinoid induced activation of apamin sensitive inhibitory neurones.²⁶ However, (+)-WIN55212 induced inhibition of NANC contractions was not modified by apamin. These NANC contractions were most likely produced by activation of postganglionic tachykinin NK₁ and NK₂ receptors as they were markedly attenuated by the combined administration of NK₁ and NK₂ antagonists but not by hexamethonium.²⁶ On the other hand, (+)-WIN55212 induced inhibition of cholinergic and NANC contractions of circular muscle was most probably not mediated by nitric oxide or opioids as this inhibitory effect was not affected by L-NAME or naloxone.²⁶

Cannabinoids inhibit intestinal motility in the whole animal

In line with the ability of cannabinoid receptor agonists to inhibit peristalsis and electrically evoked acetylcholine release and smooth muscle contractions in isolated segments of guinea pig ileum or MPLM are several reports that passage of an orally administered non-absorbable marker through the upper digestive tract can be inhibited by cannabinoids in rats³⁹⁻⁴¹ and mice (table 2). Although cannabinoids delay gastric emptying (see below), their inhibitory effect on gastrointestinal transit seems to depend at least in part on cannabinoid induced reductions in intestinal motility as the effect has also been observed in rat experiments in which the transit marker was applied intraduodenally.³⁹ As is to be expected from the *in vitro* data, the inhibitory effect of cannabinoids on intestinal transit seems to be CB₁ receptor mediated. Thus the effect is produced in a dose related fashion by the established CB₁/CB₂ receptor agonists Δ^8 -THC, Δ^9 -THC, nabilone, cannabinal, (+)-WIN55212, CP55940, and anandamide but not by the inactive (-)-enantiomer of WIN55212 or by cannabidiol^{39, 40, 44, 45, 47-51} and the effect of some CB₁ agonists on intestinal transit has been found to be susceptible to antagonism by SR141716A but not by SR144528^{40, 41, 48-51} (see also table 2) or by naloxone.^{39, 40}

Other reported *in vivo* signs of cannabinoid induced inhibition of intestinal motility are a relaxant effect on the ileum of the anaesthetised cat observed *in situ* in response to two analogues of $\Delta^{6a,10a}$ -THC,⁵² a decrease in mouse faecal water content and in intraluminal fluid accumulation in rat small intestine induced by (+)-WIN55212,⁵⁰ an inhibitory effect on faecal output, produced for example by Δ^9 -THC in rats^{53, 54} and by (+)-WIN55212 and anandamide in mice,^{50, 55} and suppression by (+)-WIN55212 and cannabinal of croton oil induced diarrhoea as measured by

Table 2 Effect of cannabinoids on transit of an orally administered non-absorbable marker through the small intestine of fasted mice

Marker	Cannabinoid	Route and time of administration	Dose or potency	Effect on transit*	Ref
Charcoal (30 min)	Δ^9 -THC	sc immediately after marker	10 or 30 mg/kg	-34%	42 43
	Δ^9 -THC		30 mg/kg	-18%	
Charcoal (15 min)	Δ^9 -THC	po at 45 min before marker	LED=5 mg/kg	-31.5%	44
	CBN		LED=10 mg/kg	-15.8%	
Charcoal (15 min)	Δ^9 -THC	po at 45 min before marker	ED ₅₀ =20 mg/kg	Inhibition	45
	Δ^9 -THC		ED ₅₀ =13.5 mg/kg	Inhibition	
Charcoal (15 min)	Δ^9 -THC	po at 45 min before marker	10 mg/kg	-58%	46
Charcoal (15 min)	Δ^9 -THC	po at 45 min before marker	LED=5 mg/kg	-26%	47
Radioactive marker (35 min)	Δ^9 -THC	iv, po or icv	ED ₅₀ =1.3 mg/kg (iv)	Inhibition	39
	CBN	immediately before marker	ED ₅₀ =12 mg/kg (iv)	Inhibition	
	Nabilone		ED ₅₀ =1.6 mg/kg (po)	Inhibition	
	Δ^9 -THC		ED ₅₀ =31.6 µg/mouse (iv)	Inhibition	
	Δ^9 -THC		ED ₅₀ =26.3 µg/mouse (icv)	Inhibition	
Charcoal (20 min)	AEA†	sc at 60 min before marker	ED ₅₀ =0.25 mg/kg	Inhibition	48
	PEA		10 mg/kg	No effect	
Carmine (20 min)	(+)-WIN†	ip at 20 min before marker	LED=0.25 mg/kg	ca. -30%	49
Charcoal (20 min)	(+)-WIN†	ip at 20 min before marker	LED=0.3 mg/kg	-34%	50
Charcoal (20 min)	(+)-WIN†	ip at 20 min before marker	ED ₅₀ =169 nmol/mouse	Inhibition	51
	CBN†		ED ₅₀ =2760 nmol/mouse	Inhibition	
	(+)-WIN†	icv at 20 min before marker	ED ₅₀ =104 nmol/mouse	Inhibition	
Charcoal (20 min)‡	CBN†		ED ₅₀ =1829 nmol/mouse	Inhibition	51
	(+)-WIN†	ip at 20 min before marker	ED ₅₀ =68 nmol/mouse	Inhibition	
	CBN†		ED ₅₀ =1681 nmol/mouse	Inhibition	
	(+)-WIN	icv at 20 min before marker	ED ₅₀ =74 nmol/mouse	Inhibition	

Time after administration of the non-absorbable marker at which the mice were killed is shown in parentheses.

sc, subcutaneously; iv, intravenously; po, orally; icv, intracerebroventricularly.

LED, lowest effective dose investigated; AEA, anandamide; CBD, cannabidiol; CBN, cannabinol; PEA, palmitylethanolamide; WIN, WIN55212.

*Compared with vehicle control.

†Antagonised by SR141716A at 1 mg/kg sc,⁴⁸ 0.62 mg/kg ip,⁴⁹ 0.3 mg/kg ip,⁵⁰ or 16 nmol/mouse ip.⁵¹

‡Experiments performed with mice exhibiting croton oil induced diarrhoea.

accelerated passage of charcoal through mouse small intestine⁵¹ (see also table 2). The ability of (+)-WIN55212 to decrease mouse faecal water content and output, and the fluid volume of rat small intestine was opposed by SR141716A, supporting the involvement of CB₁ receptors.⁵⁰ The inhibitory effects of (+)-WIN55212 and cannabinol on croton oil induced diarrhoea in mice were also antagonised by SR141716A but not by SR144528⁵¹ (see also table 2).

In contrast with results they obtained with the opioid receptor agonist loperamide, Izzo and colleagues⁴⁰ found that rat faecal output and water content were unaffected by (+)-WIN55212 or CP55940 at doses that inhibit upper intestinal transit.⁴⁰ However, they found that onset of diarrhoea induced in rats by orally administered castor oil could be delayed by CP55940, although not by (+)-WIN55212. Thus the highest dose of CP55940 used in their experiments (1 mg/kg intraperitoneally) induced transient decreases both in the incidence of diarrhoea and in faecal water content. Neither of these effects was produced by (+)-WIN55212. It should be noted that the outcome measures used in the castor oil test presumably reflect the net activity of whole gut and that there is evidence that at least one cannabinoid (Δ^9 -THC) has greater efficacy in inhibiting small intestinal transit than large bowel transit in rats.³⁹ Whether the negative results obtained in some experiments with normal or castor oil treated rats are an indication that the inhibitory effect of cannabinoids on intestinal motility is exerted mainly on the small intestine will require further research as most cannabinoid experiments relating to intestinal motility have focused on this region of the gut.

As already discussed, SR144528 does not block cannabinoid induced inhibition of intestinal transit, making it unlikely that this effect is mediated by CB₂ receptors. Even so, it would be premature to rule out the possibility that motility of some region of the gastrointestinal tract is modulated by this receptor type. This is because Hanus and colleagues⁵⁰ have reported recently that the CB₂ selective agonist HU-308 has a marked inhibitory effect on the production of faecal pellets by mice and that this effect is readily attenuated by SR144528.

Cannabinoids delay gastric emptying in the whole animal

Shook and Burks³⁹ found that intravenous administration of Δ^9 -THC inhibited the transit of a non-absorbable orally administered radioactive marker from mouse or rat stomach to small intestine, its potency in these two species being approximately the same. In mice, Δ^9 -THC was also active after intracerebroventricular injection. Other cannabinoids that have been reported to delay gastric emptying of an orally administered marker in rats or mice are WIN55212, CP55940, cannabinol, and nabilone.^{39 41 57} There is also evidence that in rats, intravenous Δ^9 -THC can reduce both the contractile activity of stomach (and duodenum) and intragastric pressure.^{39 58} It has been proposed by Krowicki and colleagues⁵⁸ that Δ^9 -THC produces its inhibitory effects on the stomach partly by acting on the dorsal vagal complex of the hind brain to modulate vagal (parasympathetic) outflow to gastric smooth muscle. This hypothesis is based on their observations that the inhibitory effect of Δ^9 -THC on stomach motility and intragastric pressure can be abolished by bilateral vagotomy at the midcervical level and by hexamethonium but not by transection of the cervical spinal cord.⁵⁸ Δ^9 -THC may also alter gastric motility by acting directly on the vagus nerves and on the gastrointestinal myenteric plexus.⁵⁸

It is likely that cannabinoids alter gastric motility by acting through CB₁ receptors. Thus the inhibitory effects of (+)-WIN55212, CP55940, and cannabinol on gastric emptying in rats and of Δ^9 -THC on rat gastric contractile activity and intragastric pressure can be prevented by SR141716A.^{41 57 58} Moreover, SR144528 does not antagonise (+)-WIN55212 or cannabinol induced inhibition of gastric emptying in rats.⁵⁷ It has also been found that the effect of Δ^9 -THC in mice on gastric emptying (and small bowel transit) is not blocked by naloxone.³⁹ By themselves neither SR141716A nor SR144528 significantly affected gastric emptying in rats.⁵⁷ However, SR141716A was unexpectedly found to share the ability of Δ^9 -THC to reduce rat intragastric pressure.⁵⁸

In agreement with animal data, McCallum and colleagues³⁹ have found Δ^9 -THC to delay gastric emptying in

human subjects fasted for eight hours when this was monitored for two hours after ingestion of a radiolabelled solid test meal and Δ^9 -THC (10 mg/m²) or placebo was administered one hour before the meal. The trial was randomised and double blind, and was performed with nine male and four female experienced cannabis users. In contrast, Bate-man⁶⁰ found that gastric emptying, monitored by a real time ultrasound technique, was unaffected by Δ^9 -THC in seven fasted cannabis naive male volunteers that had consumed 500 ml of warm orange cordial 30 minutes after injection. The doses of Δ^9 -THC used in this study (0.5 and 1 mg intravenously) increased pulse rate and produced cannabis-like psychomotor and psychological effects.

Cannabinoids inhibit gastric acid secretion

Coruzzi and colleagues⁶¹ found that (+)-WIN55212 (0.5–2 mg/kg intravenously) markedly inhibited pentagastrin stimulated gastric acid secretion in urethane anaesthetised rats without affecting basal secretion. It is likely that this effect was mediated by CB₁ receptors. Thus no such effect was observed in response to (-)-WIN55212 or to the CB₂ selective agonist JWH-015, and (+)-WIN55212 was antagonised by SR141716A and LY320135 but not by SR144528. By themselves, neither SR141716A nor LY320135 affected either basal or pentagastrin stimulated gastric acid secretion. There has also been a report that in a rat isolated stomach preparation, histamine induced but not basal gastric acid secretion is inhibited by Δ^9 -THC at the rather high concentration of 20 μ M.⁶² This effect was essentially abolished by cyclic AMP but was unaffected by propranolol. In view of these findings it is noteworthy that Nalin and colleagues⁶³ found a link between the self reported heavy smoking of cannabis by 90 human subjects (more than two days per week) and low gastric acid output. Gastric acid production was measured at least one day after admission into a quarantined study ward in which cannabis (or alcohol) consumption was not allowed. Interestingly, it was also found in this investigation that ingestion of bacterial pathogens precipitated more voluminous diarrhoea in heavy users of cannabis than in light users. Possibly this was because the lower acid content of the stomachs of the heavy users allowed more of the ingested pathogens to survive gastric transit.⁶³ Another consequence of the inhibitory effect of cannabinoids on gastric acid secretion could be the prevention of gastric ulcer formation. Indeed, gastric ulcer formation in fasted rats measured six hours after ligation of the pylorus (Shay rat test) was found to be markedly reduced by Δ^9 -THC (100 mg/kg subcutaneously or orally) when this was administered 30 minutes before ligation.⁶⁴

The endocannabinoid system of the digestive tract is tonically active

ISOLATED GUINEA PIG ILEUM

When administered by itself at concentrations of 10 nM or above, SR141716A has been found to increase the amplitude of both electrically evoked contractions of guinea pig MPLM and cholinergic and NANC contractions elicited by electrical field stimulation of a circular smooth muscle preparation of guinea pig ileum.^{13 14 26 65} That SR141716A was acting as a cholinesterase inhibitor in these tissue preparations is unlikely as it did not potentiate contractions produced by added acetylcholine. SR141716A has also been reported to augment electrically evoked acetylcholine release from guinea pig MPLM (in the presence of physostigmine)¹⁴ and to increase both distension induced ascending enteric reflex contraction of segments of ileal circular muscle³⁸ and maximal ejection pressure in isolated strips of guinea pig whole ileum exhibiting peristalsis in response to continuous intraluminal infusion of Krebs

solution.³⁷ These effects of SR141716A on acetylcholine release, evoked contractions and peristalsis, are opposite in direction to those produced in the same assay systems by cannabinoid receptor agonists.

One possible explanation for the production of such "inverse cannabimimetic effects" is that these intestinal preparations are releasing endocannabinoid(s) onto cannabinoid receptors and that SR141716A is reducing the resulting background tone by occupying these receptors. Another possible explanation springs from evidence that SR141716A is an inverse agonist rather than a "pure" or "silent" antagonist (see Pertwee⁵). According to this explanation it is assumed that cannabinoid receptors can exist in a "precoupled" state and that inverse agonists can reduce the extent of this precoupling. Consistent with the first of these hypotheses is evidence that the guinea pig ileum contains the anandamide and 2-AG metabolising enzyme FAAH, and also a mechanism for endocannabinoid uptake. Thus the presence of FAAH has been demonstrated in rat intestine⁶⁶⁻⁶⁸ and there are reports that certain effects of anandamide on guinea pig MPLM can be potentiated by the anandamide uptake inhibitor AM404, and by a general protease inhibitor phenylmethylsulphonyl fluoride (PMSF) that is known to inhibit FAAH. More specifically, experiments with guinea pig MPLM have shown PMSF to potentiate anandamide induced inhibition of electrically evoked contractions¹⁷ and AM404 to potentiate anandamide induced inhibition of resiniferatoxin evoked acetylcholine release.³⁴ However, these data should be interpreted with caution as there is now evidence that AM404 shares the ability of resiniferatoxin to activate vanilloid receptors.^{69 70} It is also noteworthy that the endocannabinoid 2-AG was first isolated from canine small intestine.⁷¹

While there is evidence for the presence in guinea pig ileum of anandamide uptake and metabolising mechanisms, there is none that anandamide is released when this tissue is electrically stimulated. Thus not all ligands capable of antagonising CB₁ mediated inhibition of electrically evoked contractions of guinea pig MPLM share the ability of SR141716A to enhance such contractions when administered alone^{23 65 72} and, when added by itself, PMSF does not mimic the inhibitory effect of anandamide on electrically evoked contractions of guinea pig MPLM.¹⁷ Moreover, while the endocannabinoid 2-AG has been detected in the small intestine of dog, there is no evidence for the presence of anandamide in the digestive tract.⁷¹ It seems likely therefore that the ability of SR141716A to produce inverse cannabimimetic effects in guinea pig ileum depends primarily on the putative inverse agonist properties of this ligand and reflects the presence in this tissue of CB₁ receptors in a precoupled state.

Although SR141716A enhances electrically evoked contractions of guinea pig MPLM, no such effect has been observed in human MPLM.¹⁸ Nor has SR141716A been found to produce effects opposite in direction to those of cannabinoid receptor agonists on threshold pressure or volume for triggering peristalsis or on longitudinal muscle reflex contraction, intestinal wall compliance, or propulsive motility measured during peristalsis in segments of guinea pig isolated ileum.^{37 38}

INTESTINAL MOTILITY IN THE WHOLE ANIMAL

Administration of SR141716A by itself, sometimes at doses slightly higher than those sufficient to antagonise cannabinoid receptor agonists, has been found to stimulate propulsion of non-absorbable markers through rat⁴⁰ or mouse intestine (table 3). That this is a reflection of the inverse agonist properties of SR141716A rather than of ongoing release of an endogenous cannabinoid receptor

Table 3 Effect of SR141716A on transit of an orally administered non-absorbable marker through the small intestine of fasted mice

Marker	Route and time of administration	Dose or potency	Effect on transit*	Ref
Charcoal (20 min)	sc before marker	1 mg/kg	+17.5%	48
Carmines (20 min)	ip at 20 min	LED=1.25 mg/kg	ca.	49
Charcoal (20 min)	before marker		+33%	
Charcoal (20 min)	ip at 20 min	LED=1 mg/kg	+48%	50
Charcoal (20 min)	before marker		Increase	
Charcoal (20 min)	ip at 20 min	ED ₅₀ =375	Increase	51
Charcoal (20 min)	before marker	nmol/mouse		
Charcoal (20 min)	icv at 20 min	ED ₅₀ =117	Increase	
Charcoal (20 min)	before marker	nmol/mouse		
Charcoal (20 min)	ip at 20 min	ED ₅₀ =418	Increase	51
(croton oil treated mice)	before marker	nmol/mouse		

Time after administration of the non-absorbable marker at which the mice were killed is shown in parentheses.

*Compared with vehicle control.

LED, lowest effective dose investigated.

agonist is suggested by data obtained *in vitro* (see above) and also by the observation that the anandamide uptake inhibitor AM404 does not affect passage of a charcoal meal along mouse intestine when administered alone or potentiate the inhibitory effect on charcoal meal transit of exogenously administered anandamide.⁴⁸ It should be borne in mind however that AM404 is not only an anandamide uptake inhibitor but also a vanilloid receptor agonist and may therefore induce acetylcholine release from the small intestine (see above).

SR141716A has also been found to increase charcoal transit through the small intestine of mice exhibiting diarrhoea induced by croton oil, albeit to no greater an extent than in croton oil free mice⁵¹ (see also table 3). In addition, it has been shown to stimulate rat or mouse defecation in some^{40 50 73} but not all experiments⁵⁵ and to increase mouse faecal water content and the fluid content of rat small intestine.^{50 73} No effect on intestinal transit has been observed in response to SR144528.^{40 51}

Central sites of action

There is some evidence that cannabinoid receptor agonists and SR141716A can affect gastrointestinal motility by interacting with sites in the brain. More specifically, there are reports that rat and mouse gastric emptying and intestinal transit can be altered by intracerebroventricular administration of Δ^9 -THC, cannabimimetic (+)-WIN55212, or SR141716A and that the changes in intestinal transit so produced can be abolished by intraperitoneal hexamethonium.^{39 41 51} However, these central sites probably contribute relatively little to the effects of peripherally administered cannabinoids as it has been found that intracerebroventricular treatment with SR141716A antagonising the inhibitory effects on gastric and intestinal transit of intracerebroventricular (+)-WIN55212 produces no such antagonism when these inhibitory effects are provoked by intraperitoneal administration of (+)-WIN55212.⁴¹ Further evidence for the relatively greater importance of peripheral sites of action comes from experiments showing that the ability of intraperitoneal SR141716A to increase mouse intestinal transit and the fluid content of rat small intestine can be attenuated or abolished by atropine but not by hexamethonium or by combined administration of NK₁ and NK₂ antagonists.⁵⁰ These findings point to the involvement of a tachykinin independent mechanism that relies on activation of peripheral muscarinic cholinergic receptors.⁵⁰ It is also likely that (+)-WIN55212 or cannabimimetic inhibit croton oil induced diarrhoea by acting peripherally, as this inhibition is also not blocked by hexamethonium.⁵¹

Gastrointestinal signs of tolerance and dependence

There are several reports that *in vivo* or *in vitro* pretreatment with cannabinoids can produce tolerance to

the inhibitory effects of these drugs on gastrointestinal activity. More specifically, Anderson and colleagues⁴⁶ found that mice pretreated with Δ^9 -THC 10 mg/kg orally once daily for 2–4 days developed tolerance to the inhibitory effect of this cannabinoid on the gastrointestinal passage of a charcoal meal. Some degree of tolerance to a challenging injection of Δ^9 -THC was still detectable even after 19 drug free days. Pertwee and colleagues⁷⁴ pretreated mice with Δ^9 -THC 20 mg/kg subcutaneously once daily for three days, or with its vehicle Tween 80. They found that the inhibitory effect of 100 nM CP55940 on electrically evoked contractions of segments of MPLM was less in tissue that had been obtained from Δ^9 -THC pretreated mice 24–28 hours after the final injection than in tissue obtained from vehicle pretreated animals. Similar results were found in guinea pigs pretreated with Δ^9 -THC 10 mg/kg intraperitoneally once daily for two days, or with its vehicle Tween 80.⁷⁵ In these experiments, Δ^9 -THC pretreatment was found to produce significant dextral shifts in the log concentration-response curves of Δ^9 -THC and CP55940 for inhibition of electrically evoked contractions of MPLM, without inducing tolerance to the contractile effect of acetylcholine. It also decreased the size of the maximal responses to Δ^9 -THC and CP55940, a possible indicator of a reduction in cannabinoid receptor density and/or coupling efficiency. Basilico and colleagues⁷⁶ exposed segments of guinea pig MPLM for five hours to a concentration of (+)-WIN55212 (50 nM) expected to reduce the amplitude of electrically evoked contractions by 50%. At the end of this period, the amplitude of evoked contractions was no less in the (+)-WIN55212 treated tissues than in untreated tissues, indicating the development of tolerance. Guagnini and colleagues⁷⁷ preincubated MPLM of human ileum or distal jejunum at 18°C for 48 hours with 10 μ M (+)- or (-)-WIN55212. Preincubation with the (+)-enantiomer but not with (-)-WIN55212 completely abolished the inhibitory effect of (+)-WIN55212 on electrically evoked contractions of MPLM. They also found that twitch responses were markedly enhanced by 1 μ M SR141716A in tissues that had been preincubated with (+)-WIN55212 but not in tissues that had been preincubated with (-)-WIN55212 or with the drug vehicle DMSO (SR141716A does not induce inverse cannabimimetic effects in human MPLM).¹⁸ Accordingly, this preparation may serve as an *in vitro* model for cannabinoid dependence. In line with the observation of Guagnini and colleagues⁷⁷ is a finding by Lichtman and colleagues⁷⁸ that one of the withdrawal signs induced by SR141716A in Δ^9 -THC tolerant dogs is diarrhoea. It is noteworthy however that SR141716A (1 μ M) has been reported not to induce “withdrawal” contractions in resting (+)-WIN55212 tolerant guinea pig MPLM.⁷⁶

In their experiments with guinea pig MPLM, Basilico and colleagues⁷⁶ also obtained evidence for the development of cross tolerance between (+)-WIN55212 and morphine. They observed significant dextral shifts in the log concentration-response curve for inhibition of electrically evoked contractions of both (+)-WIN55212 and morphine in MPLM that had been preincubated for five hours with either of these agonists. In contrast, tolerance to the inhibitory effect of normorphine (or clonidine) on electrically evoked contractions was not detected by Pertwee and colleagues⁷⁵ in guinea pig MPLM that had been rendered cannabinoid tolerant by *in vivo* pretreatment with Δ^9 -THC.

Finally, there is evidence that cannabinoid receptor agonists can suppress increases in gastrointestinal activity precipitated by naloxone in morphine dependent animals. Hine and colleagues⁷⁹ found that Δ^9 -THC but not cannabidiol produced a dose related blockade of naloxone

induced signs of heightened gastrointestinal activity (diarrhoea and increased defecation) as well as other abstinence signs in morphine dependent rats. These findings led them to conclude that cannabinoids might have potential for the management of opioid withdrawal in the clinic. Frederickson and colleagues⁸⁰ found that "withdrawal" contractions induced by naloxone in ileum taken from guinea pigs that had been treated with a slow release morphine suspension could be attenuated by (-)- but not by (+)- Δ^9 -THC at a concentration (95 nM) that did not affect contractions produced by acetylcholine. Morrone and colleagues⁸¹ reported that a cannabis extract (5.2 μ M Δ^9 -THC) reduced naloxone induced "withdrawal" contractions in segments of guinea pig ileum and rabbit jejunum that had been pre-incubated for five minutes with morphine or with the κ opioid receptor agonist U-50,488H. Acetylcholine induced contractions were not blocked by the cannabis. Basilico and colleagues⁷⁶ found that "withdrawal" contractions induced by naloxone in strips of guinea pig MPLM that had been rendered opioid and cannabinoid tolerant by a five hour exposure to morphine could be prevented by 50 nM (+)-WIN55212. Because CB₁ receptors can mediate suppression of acetylcholine release in the myenteric plexus (see above) and because naloxone precipitated "withdrawal" contractions of MPLM are thought to be triggered by the endogenous release of acetylcholine, 5-HT, and substance P into neuromuscular synapses, it is possible that (+)-WIN55212 attenuates these contractions by inhibiting the release of one or more of these contractile agents⁷⁶⁻⁸⁰ (see also above). Another possibility is that (+)-WIN55212 interacts directly with μ opioid receptors.⁷⁶

Conclusions

There is little doubt from the animal data described in this review that the endocannabinoid system extends into the enteric nervous system where it exerts an inhibitory influence on the processes of gastric emptying and peristalsis. These effects on gastrointestinal motility seem to be largely mediated by peripheral CB₁ receptors that inhibit ongoing contractile transmitter release when activated. Gastric acid secretion can also be inhibited in response to CB₁ receptor activation although the underlying mechanisms have yet to be elucidated in detail. Cannabinoid pretreatment induces tolerance to the inhibitory effects of cannabinoid receptor agonists on gastrointestinal motility. However, the ability of naloxone to induce "withdrawal" contractions in opioid pretreated intestinal tissue seems not to be shared by the CB₁ selective antagonist SR141716A when this is added to intestinal tissue that has been rendered cannabinoid tolerant by in vitro pretreatment with the cannabinoid receptor agonist (+)-WIN55212. Non-CB₁ receptors for cannabinoids that are present in the gastrointestinal tract include (i) CB₂ or CB₂-like receptors and (ii) vanilloid VR1 receptors that can be activated by anandamide and some of its analogues but not by non-icosanoid cannabinoids. However, the part played by vanilloid VR1 receptors in the pharmacology of exogenously administered or endogenously released anandamide and the role(s) of gastrointestinal CB₂ or CB₂-like receptors in health or disease remain to be established.

The observation that inverse cannabimimetic effects can be produced by the CB₁ receptor inverse agonist SR141716A in mouse, rat, or guinea pig intestine constitutes evidence that the endocannabinoid system is tonically active in the intestinal tract. This tonic activity seems to arise from the presence of a population of CB₁ receptors that are precoupled to their effector mechanisms rather than from the endogenous release of endocannabinoids. Evidence for the presence of precoupled CB₁ receptors in the human intestine has also been obtained, albeit

only in tissue (ileum) that has been rendered cannabinoid tolerant by prior exposure to a cannabinoid receptor agonist. Further experiments are now required to determine whether there are any disease states in which CB₁ receptor precoupling is increased and also whether endogenously released endocannabinoids ever contribute to the control of gastrointestinal function.

Cannabinoid receptor agonists delay gastric emptying in humans as well as in rodents, and they may also inhibit human gastric acid secretion. It is also worth noting that there have been a number of anecdotal accounts of the effective use of cannabis in the past against dysentery and cholera.⁸² Even so, the extent to which the inhibitory effects of cannabinoid receptor agonists or antagonists/inverse agonists on gastrointestinal motility and/or on gastric acid secretion can be exploited in the present day clinic has yet to be investigated in depth. So too has the extent to which these drugs provoke unwanted effects in the gastrointestinal tract when used for other medicinal purposes. The therapeutic implications of the existence of a group of drugs, the cannabinoids, that possesses anti-inflammatory and analgesic properties⁸³ coupled to an inhibitory effect on gastric acid secretion also warrants investigation. Clearly there is now a need both for clinical studies and for a more detailed elucidation through non-clinical research of the role of the endocannabinoid system in the gastrointestinal tract.

R G PERTWEE

Department of Biomedical Sciences,
Institute of Medical Sciences, University of Aberdeen
Foresterhill, Aberdeen AB25 2ZD, UK
rgp@aberd.ac.uk

Some of the research described in this article was supported by grants from the National Institute on Drug Abuse (DA 09158 and DA 09789) and from the Wellcome Trust.

- Pertwee RG. The central neuropharmacology of psychotropic cannabinoids. *Pharmacol Ther* 1988;36:189-261.
- Pertwee RG. Pharmacology of cannabinoid CB₁ and CB₂ receptors. *Pharmacol Ther* 1997;74:129-80.
- Di Marzo V, Melck D, Bisogno T, et al. Endocannabinoids: endogenous cannabinoid receptor ligands with neuromodulatory action. *Trends Neurosci* 1998;21:521-8.
- Piomelli D, Beltramo M, Glasnapp S, et al. Structural determinants for recognition and translocation by the anandamide transporter. *Proc Natl Acad Sci USA* 1999;96:5802-7.
- Pertwee RG. Pharmacology of cannabinoid receptor ligands. *Curr Med Chem* 1999;6:635-64.
- Pertwee RG. Cannabinoid receptor ligands: clinical and neuropharmacological considerations relevant to future drug discovery and development. *Exp Opin Invest Drugs* 2000;9:1553-71.
- Gill EW, Paton WDM, Pertwee RG. Preliminary experiments in the chemistry and pharmacology of cannabis. *Nature* 1970;228:134-6.
- Gill EW, Paton WDM. Pharmacological experiments in vitro on the active principles of cannabis. In: Joyce CRB, Curry SH, eds. *The botany and chemistry of cannabis*. London: Churchill, 1970:165-73.
- Rosell S, Agurell S, Martin B. Effects of cannabinoids on isolated smooth muscle preparations. In: Nahas GG, ed. *Marijuana: chemistry, biochemistry, and cellular effects*. New York: Springer-Verlag, 1976:397-406.
- Nye JS, Seltzman HH, Pitt CG, et al. High affinity cannabinoid binding sites in brain membranes labeled with [³H]-5'-trimethylammonium Δ^9 -tetrahydrocannabinol. *J Pharmacol Exp Ther* 1985;234:784-91.
- Roth SH. Stereospecific presynaptic inhibitory effect of Δ^9 -tetrahydrocannabinol on cholinergic transmission in the myenteric plexus of the guinea pig. *Can J Physiol Pharmacol* 1978;56:968-75.
- Pertwee RG, Stevenson LA, Elrick DB, et al. Inhibitory effects of certain enantiomeric cannabinoids in the mouse vas deferens and the myenteric plexus preparation of guinea pig small intestine. *Br J Pharmacol* 1992;105:980-4.
- Pertwee RG, Fernando SR, Nash JE, et al. Further evidence for the presence of cannabinoid CB₁ receptors in guinea pig small intestine. *Br J Pharmacol* 1996;118:2199-205.
- Coutts AA, Pertwee RG. Inhibition by cannabinoid receptor agonists of acetylcholine release from the guinea pig myenteric plexus. *Br J Pharmacol* 1997;121:1557-66.
- Pertwee RG, Stevenson LA, Fernando SR, et al. In vitro effects of the cannabinoid, CP55,940, and of its (+)-enantiomer, CP56,667. In: Harris L, ed. *Proceeding of the 54th annual scientific meeting of the college on problems of drug dependence*. Rockville, Maryland: National Institute on Drug Abuse, 1993:132;374.
- Mechoulam R, Devane WA, Glaser R. Cannabinoid geometry and biological activity. In: Murphy L, Bartke A, eds. *Marijuana/cannabinoids neurobiology and neurophysiology*. Boca Raton: CRC Press Inc., 1992:1-33.
- Pertwee RG, Fernando SR, Griffin G, et al. Effect of phenylmethylsulphonyl fluoride on the potency of anandamide as an inhibitor of electrically evoked contractions in two isolated tissue preparations. *Eur J Pharmacol* 1995;272:73-8.

- 18 Croci T, Manara L, Aureggi G, *et al.* *In vitro* functional evidence of neuronal cannabinoid CB₁ receptors in human ileum. *Br J Pharmacol* 1998;125:1393–5.
- 19 Rosell S, Björkroth U, Agurell S, *et al.* Relation between effects of cannabinoid derivatives on the twitch response of the isolated guinea pig ileum and their psychotropic properties. In: Nahas GG, Paton WDM, eds. *Marihuana: biological effects*. Oxford: Pergamon Press, 1979:63–70.
- 20 Lynn AB, Herkenham M. Localization of cannabinoid receptors and non-saturable high-density cannabinoid binding sites in peripheral tissues of the rat: implications for receptor-mediated immune modulation by cannabinoids. *J Pharmacol Exp Ther* 1994;268:1612–23.
- 21 Anavi-Goffer S, Irving A, Pertwee R, *et al.* The cellular distribution of CB₁ receptors in the myenteric plexus. *International Cannabinoid Research Society 2000 Symposium on the Cannabinoids*. Burlington, Vermont, 2000:1.
- 22 Buckley NE, Hansson S, Harta G, *et al.* Expression of the CB₁ and CB₂ receptor messenger RNAs during embryonic development in the rat. *Neuroscience* 1998;82:1131–49.
- 23 Ross RA, Brockie HC, Fernando SR, *et al.* Comparison of cannabinoid binding sites in guinea pig forebrain and small intestine. *Br J Pharmacol* 1998;125:1345–51.
- 24 Griffin G, Fernando SR, Ross RA, *et al.* Evidence for the presence of CB₂-like cannabinoid receptors on peripheral nerve terminals. *Eur J Pharmacol* 1997;339:53–61.
- 25 Shire D, Carillon C, Kaghad M, *et al.* An amino-terminal variant of the central cannabinoid receptor resulting from alternative splicing. *J Biol Chem* 1995;270:3726–31.
- 26 Izzo AA, Mascolo N, Borrelli F, *et al.* Excitatory transmission to the circular muscle of the guinea pig ileum: evidence for the involvement of cannabinoid CB₁ receptors. *Br J Pharmacol* 1998;124:1363–8.
- 27 Rinaldi-Carmona M, Barth F, Héaulme M, *et al.* SR141716A, a potent and selective antagonist of the brain cannabinoid receptor. *FEBS Letters* 1994;350:240–4.
- 28 Coutts AA, Pertwee RG. Evidence that cannabinoid-induced inhibition of electrically evoked contractions of the myenteric plexus-longitudinal muscle preparation of guinea pig small intestine can be modulated by Ca²⁺ and cAMP. *Can J Physiol Pharmacol* 1998;76:340–6.
- 29 Rosell S, Agurell S. Effects of 7-hydroxy- Δ^9 -tetrahydrocannabinol and some related cannabinoids on the guinea pig isolated ileum. *Acta Physiol Scand* 1975;94:142–4.
- 30 Ward SJ, Mastriani D, Casiano F, *et al.* Pravadinol: profile in isolated tissue preparations. *J Pharmacol Exp Ther* 1990;255:1230–9.
- 31 Pertwee RG, Browne SE, Ross TM, *et al.* An investigation of the involvement of GABA in certain pharmacological effects of delta-9-tetrahydrocannabinol. *Pharmacol Biochem Behav* 1991;40:581–5.
- 32 Layman JM, Milton AS. Some actions of Δ^1 -tetrahydrocannabinol and cannabidiol at cholinergic junctions. *Br J Pharmacol* 1971;41:379–80P.
- 33 Paton WDM, Pertwee RG, Temple D. The general pharmacology of cannabinoids. In: Paton WDM, Crown J, eds. *Cannabis and its derivatives*. Oxford: Oxford University Press, 1972:50–75.
- 34 Yal WM, Reese LA. Cannabinoid receptor: inhibition of ACh release from myenteric plexus of small intestine. *Gastroenterology* 1999;116:G4791.
- 35 López-Redondo F, Lees GM, Pertwee RG. Effects of cannabinoid receptor ligands on electrophysiological properties of myenteric neurones of the guinea pig ileum. *Br J Pharmacol* 1997;122:330–4.
- 36 Fernando SR. *Development of in vitro smooth muscle preparations as suitable models for elucidating the mechanism of action of cannabinoids*, PhD Thesis. Aberdeen: University of Aberdeen, 1998.
- 37 Izzo AA, Mascolo N, Tonini M, *et al.* Modulation of peristalsis by cannabinoid CB₁ ligands in the isolated guinea pig ileum. *Br J Pharmacol* 2000;129:984–90.
- 38 Heinemann A, Shahbazian A, Holzer P. Cannabinoid inhibition of guinea pig intestinal peristalsis via inhibition of excitatory and activation of inhibitory neural pathways. *Neuropharmacology* 1999;38:1289–97.
- 39 Shook JE, Burks TF. Psychoactive cannabinoids reduce gastrointestinal propulsion and motility in rodents. *J Pharmacol Exp Ther* 1989;249:444–9.
- 40 Izzo AA, Mascolo N, Pinto L, *et al.* The role of cannabinoid receptors in intestinal motility, defaecation and diarrhoea in rats. *Eur J Pharmacol* 1999;384:37–42.
- 41 Manara L, Landi M, Croci T, *et al.* Delay of gastric emptying and gastrointestinal transit by stimulation of intestinal cannabinoid CB₁ receptors. *FASEB J* 2000;14:785.
- 42 Dewey WL, Kennedy JS, Howes JF. Some autonomic, gastrointestinal and metabolic effects of two constituents of marihuana. *Fed Proc* 1970;29:650.
- 43 Dewey WL, Harris LS, Kennedy JS. Some pharmacological and toxicological effects of l-trans- Δ^8 - and l-trans- Δ^9 -tetrahydrocannabinol in laboratory rodents. *Arch Int Pharmacodyn* 1972;196:133–45.
- 44 Anderson PF, Jackson DM, Cheshier GB. Interaction of Δ^9 -tetrahydrocannabinol and cannabidiol on intestinal motility in mice. *J Pharm Pharmacol* 1974;26:136–7.
- 45 Cheshier GB, Dahl CJ, Everingham M, *et al.* The effect of cannabinoids on intestinal motility and their antinociceptive effect in mice. *Br J Pharmacol* 1973;49:588–94.
- 46 Anderson PF, Jackson DM, Cheshier GB, *et al.* Tolerance to the effects of Δ^9 -tetrahydrocannabinol in mice on intestinal motility, temperature and locomotor activity. *Psychopharmacologia* 1975;43:31–6.
- 47 Jackson DM, Malor R, Cheshier GB, *et al.* The interaction between prostaglandin E₁ and Δ^9 -tetrahydrocannabinol on intestinal motility and on the abdominal constriction response in the mouse. *Psychopharmacology* 1976;47:187–93.
- 48 Calignano A, La Rana G, Makriyannis A, *et al.* Inhibition of intestinal motility by anandamide, an endogenous cannabinoid. *Eur J Pharmacol* 1997;340:R7–8.
- 49 Colombo G, Agabio R, Lobina C, *et al.* Cannabinoid modulation of intestinal propulsion in mice. *Eur J Pharmacol* 1998;344:67–9.
- 50 Izzo AA, Mascolo N, Borrelli F, *et al.* Defaecation, intestinal fluid accumulation and motility in rodents: implications of cannabinoid CB₁ receptors. *Naunyn Schmiedeberg Arch Pharmacol* 1999;359:65–70.
- 51 Izzo AA, Pinto L, Borrelli F, *et al.* Central and peripheral cannabinoid modulation of gastrointestinal transit in physiological states or during the diarrhoea induced by croton oil. *Br J Pharmacol* 2000;129:1627–32.
- 52 Dagirmanjian R, Boyd ES. Some pharmacological effects of two tetrahydrocannabinols. *J Pharmacol Exp Ther* 1962;135:25–33.
- 53 Masur J, März RMW, Carlini EA. Effects of acute and chronic administration of Cannabis sativa and (-)- Δ^9 -trans-tetrahydrocannabinol on the behaviour of rats in an open-field arena. *Psychopharmacologia* 1971;19:388–97.
- 54 Drew WG, Miller LL, Wikler A. Effect of Δ^9 -THC on the open-field activity of the rat. *Psychopharmacologia* 1972;23:289–99.
- 55 Costa B, Vailati S, Colleoni M. SR 141716A, a cannabinoid receptor antagonist, reverses the behavioural effects of anandamide-treated rats. *Behav Pharmacol* 1999;10:327–31.
- 56 Hanus L, Breuer A, Tchilibon S, *et al.* HU-308: A specific agonist for CB₂, a peripheral cannabinoid receptor. *Proc Natl Acad Sci USA* 1999;96:14228–33.
- 57 Izzo AA, Mascolo N, Capasso R, *et al.* Inhibitory effect of cannabinoid agonists on gastric emptying in the rat. *Naunyn Schmiedeberg Arch Pharmacol* 1999;360:221–3.
- 58 Krowicki ZK, Moerschbaecher JM, Winsauer PJ, *et al.* Δ^9 -tetrahydrocannabinol inhibits gastric motility in the rat through cannabinoid CB₁ receptors. *Eur J Pharmacol* 1999;371:187–96.
- 59 McCallum RW, Soykan I, Sridhar KR, *et al.* Delta-9-tetrahydrocannabinol delays the gastric emptying of solid food in humans: a double-blind, randomized study. *Aliment Pharmacol Ther* 1999;13:77–80.
- 60 Bateman DN. Delta-9-tetrahydrocannabinol and gastric emptying. *Br J Clin Pharmacol* 1983;15:749–51.
- 61 Coruzzi G, Adami M, Coppelli G, *et al.* Inhibitory effect of the cannabinoid receptor agonist WIN 55,212-2 on pentagastrin-induced gastric acid secretion in the anaesthetized rat. *Naunyn Schmiedeberg Arch Pharmacol* 1999;360:715–18.
- 62 Rivas-V JF, Garcia R. Inhibition of histamine-stimulated gastric acid secretion by Δ^9 -tetrahydrocannabinol in rat isolated stomach. *Eur J Pharmacol* 1980;65:317–18.
- 63 Nalin DR, Rhead J, Rennels M, *et al.* Cannabis, hypochlorhydria and cholera. *Lancet* 1978;2:859–62.
- 64 Sofia RD, Diamantis W, Harrison JE, *et al.* Evaluation of antiulcer activity of Δ^9 -tetrahydrocannabinol in the Shay rat test. *Pharmacology* 1978;17:173–7.
- 65 Coutts AA, Brewster N, Ingram T, *et al.* Comparison of novel cannabinoid partial agonists and SR141716A in the guinea pig small intestine. *Br J Pharmacol* 2000;129:645–52.
- 66 Desarnaud F, Cadas H, Piomelli D. Anandamide amidohydrolase activity in rat brain microsomes. Identification and partial characterization. *J Biol Chem* 1995;270:6030–5.
- 67 Katayama K, Ueda N, Kurahashi Y, *et al.* Distribution of anandamide amidohydrolase in rat tissues with special reference to small intestine. *Biochim Biophys Acta* 1997;1347:212–18.
- 68 Ueda N, Yamamoto S. Anandamide amidohydrolase (fatty acid amide hydrolase). *Prostaglandins Other Lipid Mediators* 2000;61:19–28.
- 69 Jerman JC, Brough SJ, Davis JB, *et al.* The anandamide transport inhibitor AM404 is an agonist at the rat vanilloid receptor (VR1). *Br J Pharmacol* 2000;129:73P.
- 70 Zygmunt PM, Chuang HH, Movahed P, *et al.* The anandamide transport inhibitor AM404 activates vanilloid receptors. *Eur J Pharmacol* 2000;396:39–42.
- 71 Mechoulam R, Ben-Shabat S, Hanus L, *et al.* Identification of an endogenous 2-monoacylglyceride, present in canine gut, that binds to cannabinoid receptors. *Biochem Pharmacol* 1995;50:83–90.
- 72 Coutts A, Pertwee R. A comparison of cannabinoid CB₁ antagonism and inverse agonism in guinea pig myenteric neurones. *International Cannabinoid Research Society 2000 Symposium on the Cannabinoids*. Burlington, Vermont, 2000:46.
- 73 Costa B, Colleoni M. SR141716A induces in rats a behavioral pattern opposite to that of CB₁ receptor agonists. *Acta Pharmacologica Sinica* 1999;20:1103–8.
- 74 Pertwee RG, Stevenson LA, Fernando SR, *et al.* The production of cannabinoid tolerance in preparations of mouse vas deferens and of myenteric plexus-longitudinal muscle of mouse small intestine. *Br J Pharmacol* 1992;107:365P.
- 75 Pertwee R, Fernando S, Ritchie J, *et al.* Preliminary validation of a novel experimental model for the study of cannabinoid tolerance. *International Cannabinoid Research Society 1998 Symposium on the Cannabinoids*. Burlington, Vermont, 1998:127.
- 76 Basilio L, Parolaro D, Colleoni M, *et al.* Cross-tolerance and convergent dependence between morphine and cannabimimetic agent WIN 55,212-2 in the guinea pig ileum myenteric plexus. *Eur J Pharmacol* 1999;376:265–71.
- 77 Guagnini F, Croci T, Aureggi G, *et al.* Tolerance to (+)-WIN55212-2 inhibitory effect and withdrawal by the cannabinoid CB₁ receptor antagonist SR141716 in isolated strips of human small intestine. *International Cannabinoid Research Society 2000 Symposium on the Cannabinoids*. Burlington, Vermont, 2000:82.
- 78 Lichtman AH, Wiley JL, LaVecchia KL, *et al.* Effects of SR 141716A after acute or chronic cannabinoid administration in dogs. *Eur J Pharmacol* 1998;357:139–48.
- 79 Hine B, Friedman E, Torrelío M, *et al.* Morphine-dependent rats: blockade of precipitated abstinence by tetrahydrocannabinol. *Science* 1975;187:443–5.
- 80 Frederickson RCA, Hewes CR, Aiken JW. Correlation between the *in vivo* and *in vitro* expression of opiate withdrawal precipitated by naloxone: their antagonism by l-(-)- Δ^9 -tetrahydrocannabinol. *J Pharmacol Exp Ther* 1976;199:375–84.
- 81 Morrone LA, Romanelli L, Mazzanti G, *et al.* Hashish antagonism on the *in vitro* development of withdrawal contracture. *Pharmacol Res* 1993;27(suppl 1):63–4.
- 82 Mechoulam R. The pharmacology of Cannabis sativa. In: Mechoulam R, ed. *Cannabinoids as therapeutic agents*. Boca Raton: CRC Press Inc., 1986:1–19.
- 83 Pertwee RG. Cannabinoid receptors and pain. *Prog Neurobiol* (in press).